Possible vertical transmission of *Babesia canis canis* from a bitch to her puppies: a case report

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**ABSTRACT:** The present study reports the possible vertical transmission of *Babesia canis canis* from an infected bitch to her puppies. The study concerns a bitch that had developed babesiosis in week seven of pregnancy and her litter, three puppies that exhibited symptoms of the disease in Weeks 8–9 post-partum. In all animals, the infection with protozoa was confirmed by the results of a PCR blood test. The identity of the nucleotide sequences of the amplified fragment of the gene (18S RNA) isolated from the blood of the puppies and the bitch was 100%, which indicates that all the dogs were infected with the same strain of protozoa. This result, together with the exclusion of other possible routes of babesiosis transmission in puppies, suggests that they were infected with *Babesia canis canis in utero.*

**Keywords:** *Babesia canis canis*; dogs; vertical transmission

Canine babesiosis is a common and clinically significant tick-borne disease caused by haematozoan parasites of the genus *Babesia* (Adaszek and Winiarczyk 2008). The classification of *Babesia* spp. places them in the order Piroplasmida within the phylum Apicomplexa. Two morphologically distinct forms of the erythrocytic stage in the canine host were recognized in early studies that led to the naming of the larger form, measuring approximately 3–5 μm as *B. canis*, and the smaller (1–3 μm) as *B. gibsoni*. On the basis of cross-immunity, serological testing, vector specificity and molecular phylogeny *Babesia canis* was reclassified into three sub-species: *B. canis canis*, *B. canis rossi*, and *B. canis vogeli*. All of them are now considered to be separate species (Zahler et al. 1998; Matijatko et al. 2012). So far, only *B. canis* has been found in dogs in Poland (Zygner et al. 2007; Adaszek et al. 2011). These parasites are also the most common etiologic factor of babesiosis in dogs in other parts of Europe (Cardoso et al. 2008; Solano-Gallego et al. 2008; Kubelova et al. 2011). Clinically, these pathogens cause remittent fever, progressive anaemia, haemoglobinuria, and marked splenomegaly and hepatomegaly in dogs and, in some cases, the death of infected animals (Milczak et al. 2004; Adaszek et al. 2009). The most common mode of babesiosis transmission is by tick bite, but also other routes of transmission such as direct transmission via blood transfer from dog bites or blood transfusion are possible. Transplacental transmission has also been described (Fukamoto et al. 2005).

The present study reports the possible vertical transmission of *Babesia canis canis* from an infected bitch to her puppies.

**Case description**

In September 2013, a four-year-old Russian Terrier bitch was admitted to the Department of Infectious Diseases, Faculty of Veterinary Medicine in Lublin, Poland. The bitch was in Week 7 of pregnancy, showing the following symptoms: apathy,
lack of appetite, fever (40.2 °C), pale mucus membranes and brown-coloured urine. About four days earlier, the owner had removed three ticks from the animal’s skin. Blood tests revealed anaemia (RBC = 3.24 × 10^{12}/l; reference ranges 5.5–8.0 × 10^{12}/l), leukopenia (WBC = 5.1 × 10^{9}/l; reference ranges 6–16.5 × 10^{9}/l), and thrombocytopenia (PLT = 62 × 10^{9}/l; reference ranges 200–580 × 10^{9}/l). Biochemical serum tests demonstrated elevated levels of AST (62 IU; reference ranges 1–37 IU), ALT (88 IU; reference ranges 3–50 IU), urea (11.14 mmol/l; reference range 3.32 to 7.47 mmol/l), total bilirubin (2.2 mg/dl; reference ranges < 0.6 mg/dl) and creatinine (187.2 µmol/l; reference ranges 88.4–150.3 µmol/l). The Giemsa-stained blood smear demonstrated the presence of Babesia protozoa within the erythrocytes. Based on the results of the PCR test performed by Adaszek and Winiarczyk (2008) and sequencing of the obtained amplification products, the protozoa were classified as the 18S RNA-B strain. The abdominal ultrasound in the bitch revealed enlargement of the spleen; all foetuses (n = 3) were alive. The animal was administered a single subcutaneous injection of imidocarb dipropionate (5 mg/kg), which resulted in a significant improvement 24 h following administration of the drug. The bitch became more vivid and regained appetite. Control haematological and biochemical tests performed one week after administration of imidocarb only demonstrated persistent mild thrombocytopenia (PLT = 154 × 10^{9}/l). On the 59th day after mating, the bitch gave birth to three healthy puppies (one female No. 1, and two males No. 2 and No. 3). In Week 7 after birth, the puppies were weaned and sold. In Week 8, in the female (No. 1) and in one male (No. 2), and in Week 9 in the other male (No. 3), the owners observed symptoms typical of babesiosis (apathy, lack of appetite, brown urine (males No. 2 and No. 3), pallor (male No. 2), yellowing of the mucous membranes (female No. 1 and male No. 3) and vomiting and diarrhoea (female No. 1).

In all animals, the blood tests revealed anaemia, thrombocytopenia and leukopenia (Table 1). The microscopic examination of blood smears demonstrated the presence of Babesia protozoa in the erythrocytes of both males (No. 2 and No. 3), whereas PCR testing showed the presence of B. canis 18S RNA-B genetic material in the blood of all three puppies. The identity of the nucleotide sequences of the amplified gene fragment isolated from the blood of the puppies and the bitch was 100%, which indicates that all animals were infected with the same strain of protozoa.

A single subcutaneous injection of imidocarb dipropionate (5 mg/kg) administered to all three puppies resulted in a significant improvement in their health status 24–72 h after drug administration. Control examination (PCR) of the blood samples collected two weeks after therapy did not reveal the presence of genetic material of Babesia canis in any of the studied puppies.

**DISCUSSION AND CONCLUSIONS**

As in all the puppies fipronil (spray) was applied to prevent ticks and none of them showed the presence of ticks on the skin, the animals did not receive (except for a prophylactic vaccination and anti-parasitic prophylaxis) any medical-veterinary treatment (iatrogenic infection could be excluded). Thus, it is assumed that the probable route of Babesia transmission was in utero infection. This is also supported by the 100% identity of DNA fragments of the 18S RNA B gene of B. canis isolated from the mother and puppies, suggesting that all four dogs were infected with the same strain of the parasite.

However, other routes of transmission of Babesia should be considered, such as congenital transmission in the birth canal or through colostrum. So far there are no data about these modes of transmission of canine babesiosis. It is documented (Perez

**Table 1. The results of haematological examination of the puppies infected with Babesia canis**

<table>
<thead>
<tr>
<th>Number of animals</th>
<th>RBC (10^{12}/l)</th>
<th>Ht (%)</th>
<th>WBC (10^9/l)</th>
<th>PLT (10^9/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. 1 (female)</td>
<td>4.2</td>
<td>33</td>
<td>5.5</td>
<td>88</td>
</tr>
<tr>
<td>No. 2 (male)</td>
<td>4.75</td>
<td>38</td>
<td>5.1</td>
<td>81</td>
</tr>
<tr>
<td>No. 3 (male)</td>
<td>4.0</td>
<td>33</td>
<td>4.9</td>
<td>100</td>
</tr>
<tr>
<td>Reference ranges</td>
<td>5.5–8</td>
<td>37–55</td>
<td>6–16.5</td>
<td>200–580</td>
</tr>
</tbody>
</table>
et al. 1996; Allsopp et al. 2007) that seropositive dams are able to transmit Babesia antibodies (but not the parasites) to their new-borns through colostrum, but these antibodies will decrease in time after birth, so this way of transmission may be excluded in the present case.

In the available literature there are very little data concerning the possible vertical transmission of B. canis canis from a bitch to her puppies. Mierzejewska et al. (2014) presented the case of vertical transmission of B. canis in a litter of Central Asian Shepherds. The DNA of the protozoa was detected using PCR in three pups with babesiosis symptoms, and in their asymptomatic mother. The isolates derived from the pups and the female – 520 bp 18S rRNA gene fragments – were compared and analysed. All isolates from the pups and their mother were identical and showed 100% identity with B. canis EU622793, supporting a common source of infection.

The transplacental transmission of B. gibsoni has been experimentally demonstrated in a bitch that delivered a litter of one stillborn and four live pups (Fukumoto et al. 2005). These four pups died of congenital babesiosis between Days 14 and 39 post-birth. For Babesia infection, there are reports describing the possibility of congenital babesiosis in horses (Neitz 1956; Allsopp et al. 2007), calves (De Vos et al. 1976; Yeruham et al. 2003), and sheep (Neitz 1956).

In horses, once an animal is infected with T. equi, it remains a lifelong carrier, since anti-theilerial drugs do not completely eliminate the parasite (Allsopp et al. 2007). Infected mares can transmit T. equi piroplasms across the placenta and this might result in abortion or neonatal piroplasmosis. Colostral antibodies to T. equi may suppress parasitaemia in new-born foals thereby reducing the incidence of clinical neonatal equine piroplasmosis, which could control parasitaemia during the early months of life (Allsopp et al. 2007). In the present report, an almost two-month time interval between possible infection in utero and clinical disease might be explained by a similar protection conferred on the pups by maternal colostrum. Nevertheless, and although they had no detectable ticks, the possibility that the pups were infected after birth by a tick vector cannot be ruled out completely (Simoes et al. 2011).

A similar case of a dog infected with B. microti-like piroplasms, probably in utero, was reported by Simoes et al. (2011). The disease occurred in a two-month old female German Shepherd bitch. As the pup had never had detectable ticks, did not travel, and came from an area where B. microti-like piroplasms are not endemic, it was concluded that the disease was the result of vertical transmission.

From a clinical standpoint, the development of a highly sensitive diagnostic method for congenital canine babesiosis is required. The detection of maternal antibodies is unsuitable because they fall below the detectable level a few weeks after birth (Fukumoto et al. 2005). Also, as shown by the results of our observations, microscopic examination has limited sensitivity: the presence of parasites in Giemsa-stained blood smears was only demonstrated in two out of the three infected pups. Only PCR could detect the parasites in all pups. Therefore, we suggest that a PCR-based test is most suitable for the specific diagnosis of congenital babesiosis.

REFERENCES


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